

Article

Catalytic and Biological Activity of Silver and Gold Complexes Stabilized by NHC with Hydroxy Derivatives on Nitrogen Atoms

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Abstract: In this paper is reported the synthesis of N,N' hydroxy derivative of NHC silver (**3a–4a**) and gold(I) (**3b–4b**) complexes of general formula $[M(NHC)_2]^+ [MX_2]^-$. All compounds were characterized by spectroscopic and analytic techniques. The complexes turned out to be effective in both catalytic and biological applications. They catalyzed the coupling of aldehyde, piperidine, and phenylacetylene in A^3 -reaction to produce propargylamines and showed antimicrobial activity. In fact, minimal inhibition concentration (MIC) tests with Gram-positive and Gram-negative bacteria demonstrated that the silver compounds are selective toward *E. coli*, whereas the gold analogues are active against *S. aureus*. Moreover, the N,N' hydroxy derivative of NHC silver complexes **3a** and **4a** exhibited good anticancer activity on the HeLa cancer cells ($3a$ -IC₅₀ = 12.2 ± 0.1 μM, $4a$ -IC₅₀ = 11.9 ± 1.2 μM), whereas gold complex **4b** displayed good anticancer activity towards the MCF-7 cells (IC₅₀ = 12.2 ± 1.2 μM).

Keywords: NHC silver and gold complexes; coupling-catalysis; cytotoxic activity



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1. Introduction

N-heterocyclic carbenes (NHCs) are nucleophilic compounds widely studied for their strong coordination ability as ancillary ligands of transition metals (M-NHCs) [1]. Their properties (steric and electronic) can be easily modulated, varying the substituents on the backbone and/or on the nitrogen atoms within the cyclic structure [1,2]. This versatility has made them suitable ligands in metal complexes, capable of catalysis [3–5] and medicinal applications [6–8].

Among the many transition elements stabilized by these ligands, coinage metals play an important role. They have proved to be useful activators in homogeneous catalysis for the hydrofunctionalization of allenes [9], alkenes [10] and alkynes [11], cycloisomerisations of enynes and ynones [12], C–H activation [13], and other organic transformations [14], as well as carbene transfer reactions [15]. Furthermore, in the past 15 years, the Ag(I) and Au(I) NHC-complexes have been studied in the catalysis of new and, until then, unexplored reactions [14,16]. NHC-Ag complexes have been tested in reactions of diboration of alkenes [17], in the 1,3-dipolar cycloadditions of azomethine ylides with prop-2-enoates [18], in the cyclopropanation reactions between styrene and phenyldiazoacetate [19] and in the Sonogashira coupling of phenylacetylene and 4-iodoacetophenone [20].

NHC-Au complexes have been studied for addition of nucleophiles (water [21,22], alcohols [23], and amines [11]) to non-activated alkynes, in the A^3 -coupling reactions [24], and in the carboxylation of propargylamines to yield 2-oxazolidinones [25]. These Au

2. Results and Discussion

2.1. Synthesis and Characterization

The NHC metal complexes, represented in Figure 2, were synthesized by slight changes of the literature procedure [37–43].

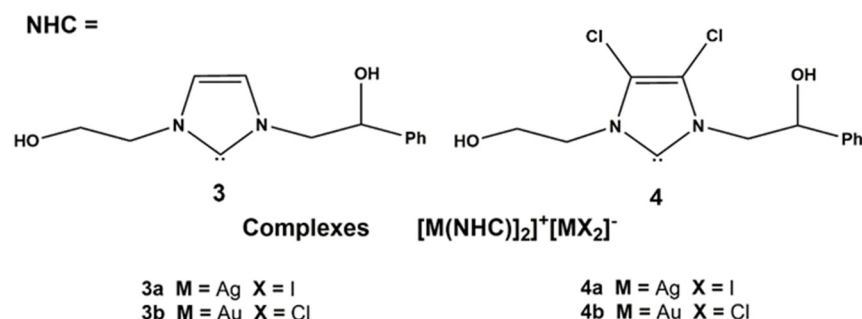
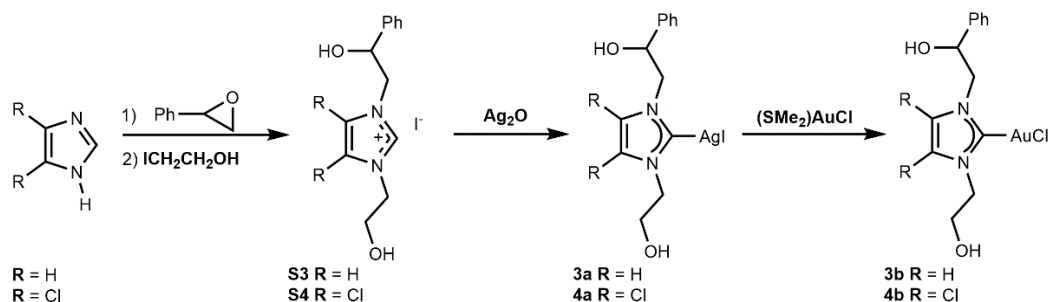


Figure 2. NHC ligands (3–4) and silver(I) (3a–4a) and gold(I) (3b–4b) complexes synthesized in this work.

The synthetic route to obtain N-heterocyclic carbene complexes **3a**, **3b**, **4a**, and **4b** is depicted in Scheme 1. The first step is the synthesis of two imidazolium salts (**S3** and **S4**) containing different substituents on the backbone (R = H for **S3** or R = Cl for **S4**). They were achieved by reaction of imidazole or 4,5-dichloroimidazole with phenylethylene oxide followed by addition of 2-iodoethanol (see Scheme 1).



Scheme 1. Synthesis of NHC silver(I) and gold(I) complexes **3a**, **3b**, **4a**, and **4b**.

The obtained salts **S3** and **S4** were characterized by ^1H - and ^{13}C -NMR analysis and by mass spectrometry. In ^1H -NMR spectra, the signals of the protons on carbocationic (NCHN protons) of **S3** and **S4** are at 9.12 ppm and 9.51 ppm, respectively. In ^{13}C NMR spectra, the resonance of NCHN carbocationics are at 136.75 ppm and 137.39 ppm for **S3** and **S4**, respectively. The mass spectra (MALDI) of **S3** show singlet signals at 233.12898, whereas **S4** show multiplet signals having the main peak at 301.05142 Dalton, attributable to $[\text{C}_{13}\text{H}_{17}\text{N}_2\text{O}_2]^+$ and $[\text{C}_{13}\text{H}_{15}\text{Cl}_2\text{N}_2\text{O}_2]^+$ that represent the cationic portions of imidazolium salts. The reaction of **S3** or **S4** salts with silver oxide (Ag_2O) produced the corresponding silver(I) complexes **3a** and **4a** (see Scheme 1). They were characterized by NMR spectroscopy, ESI-MS spectrometry, and elemental analysis. The ^1H and ^{13}C NMR spectra show all the expected signals. In the ^1H -NMR spectra, the absence of the signal attributable to the proton on NCHN carbocation demonstrated that the deprotonation reaction by silver oxide with formation of carbene has taken place [14,15]. The formation of **3a** silver complex is confirmed in the ^{13}C -NMR spectrum by the presence of a doublet attributable to $\text{C}_{\text{carbene}}\text{NCN-Ag}$. This type of signal is due to the coupling constant of $\text{C}_{\text{carbene}}$ and Ag, whose two naturally isotopes (^{107}Ag 51.839% and ^{109}Ag 48.161%) are both NMR active, having nuclear spin $\frac{1}{2}$. For **3a** the coupling constants are 182.3 Hz and 209.1 Hz, respectively. Thus, the mass spectrum of **3a** (ESI-MS) shows two signals of almost the same intensity attributable to $[\text{C}_{26}\text{H}_{32}\text{AgN}_4\text{O}_4]^+$, corresponding to $[(\text{NHC})_2\text{Ag}]^+$. The multiplicity of

signals is due to nearly equal natural abundance of two isotopes of silver (^{107}Ag and ^{109}Ag) [44]. The ^{13}C -NMR spectrum of **4a** shows all the expected signals and a sharp signal attributable to carbene carbon bonded to silver atom ($\text{C}_{\text{carbene-Ag}}$) at 183.67 ppm. It is worth noting that in literature are reported three cases for the pattern resonance of the carbene carbons bonded to silver ($\text{C}_{\text{carbene-Ag}}$), i.e., (i) doublet of doublets (coupling constants of each silver isotope ^{107}Ag and ^{109}Ag); (ii) no splitting pattern (sharp or broad singlet); and (iii) no observation of the carbenic peak [44]. The ESI-MS spectrum of **4a** shows more signals attributable to a bis-carbene structure $[\text{C}_{26}\text{H}_{28}\text{AgCl}_4\text{N}_4\text{O}_4]^+$. In this case the multiplicity of signals is attributable to the presence of both silver isotopes (^{107}Ag and ^{109}Ag) and chlorine (^{35}Cl and ^{37}Cl). The elemental analysis for two complexes shows a relationship between ligand, silver, and iodide of 1:1:1, so that it is reasonable to assume for both complexes a structure of the type $[(\text{NHC})_2\text{Ag}]^+[\text{AgI}_2]^-$. Whereby, silver complexes **3a** and **4a**, according to mass and elemental analysis, are present as $[(\text{NHC})_2\text{Ag}]^+$ cation and reasonably $[\text{AgI}_2]^-$ as counter-ion. The proposed structure was observed in solid state for similar silver complexes by X-ray diffraction analysis from both Lin [15] and by some of us [45].

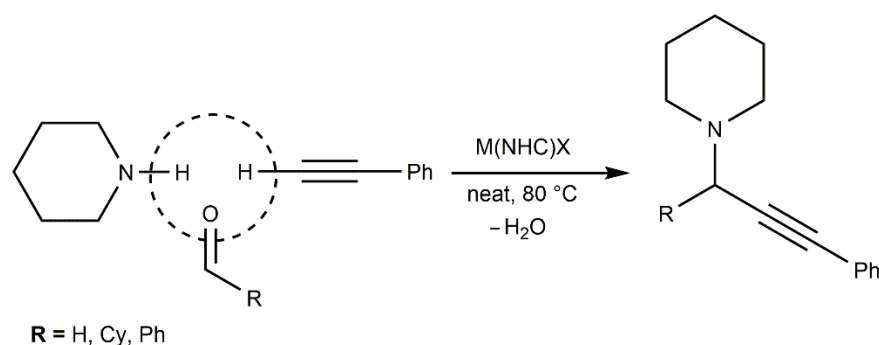
The gold complexes **3b** and **4b** were synthesized via trans-metalation (see Scheme 1) by reaction of silver complex **3a** or **4a** with dimethylsulfide-gold(I)-chloride at room temperature and with exclusion of light. The two gold complexes were obtained as light-yellow powder in a good yield: 40% for **3b** and 61% for **4b**. The complexes were characterized by ^1H -NMR, ^{13}C -NMR, mass spectrometry (MALDI-MS), and elemental analysis. The ^{13}C -NMR spectra of **3b** and **4b** show sharp signals for carbene carbon at 168.37 and 170.90 ppm, respectively. Mass spectra show a peak at 661.2110 for **3b** and 709.05720 for **4b** m/z , attributable to a bis-carbene structure. All the complexes are moisture and light stable; when solubilized in DMSO/ D_2O (90/10) and exposed to light for 24 h at room temperature, they did not show any changes in ^1H -NMR spectra. The elemental analysis for two complexes shows a relationship between ligand, gold, and chloride of 1:1:1, so that it is reasonable to assume also for gold(I) complexes a structure of the type $[(\text{NHC})_2\text{Au}]^+[\text{AuCl}_2]^-$. However, it is fair to suppose that for these complexes in solution there is an equilibrium between the ionic form $[\text{M}(\text{NHC})_2]^+[\text{MX}_2]^-$ and neutral form $\text{M}(\text{NHC})\text{X}$. Thus, conductivity measurements confirmed the electrolytic nature of the complexes; in fact, the conductance values of the compounds, measured in CH_2Cl_2 , displayed a concentration dependence. Whereas it is obvious to believe that the neutral specie is present in solution because a gold complex in neutral form was recently isolated and characterized by X-ray diffraction analysis, and this is the necessary form for catalytic activity [46].

2.2. Catalytic Behavior in A^3 -Coupling Reactions

In literature, it is known that many metal complexes can catalyze the synthesis of important scaffolds [24,32], and recently, among them, *N*-heterocyclic carbene complexes of silver and gold have showed valuable catalytic activity in A^3 -coupling reactions [37,46,47].

For this reason, the silver and gold complexes **3a**, **4a**, **3b**, and **4b** were tested in A^3 -coupling reactions of aldehyde, amine, and alkyne to produce propargylamines (Scheme 2). They are an important class of organic compound precursor of drugs that act as neuro-modulator of monoamine oxidases (MAO), which are a valuable class of mitochondrial enzymes with a critical role in the inhibitory activity of dopaminergic metabolism, i.e., pargyline, rasagyline, selegiline, that are used in the treatment of Alzheimer's and Parkinson's diseases [33–35].

In Table 1 we report the catalytic activities of the synthesized NHC silver and gold complexes using alkyne (phenylacetylene), secondary amine (piperidine), and three different aldehydes (*p*-formaldehyde, cyclohexanecarboxaldehyde, and benzaldehyde) at 80 °C to obtain PAA1, PAA2, and PAA3.



Scheme 2. A³-coupling reaction to give propargylamine PAA (PAA1 where R = H; PAA2 where R = cyclohexyl; PAA3 where R = phenyl).

Table 1. Catalytic activity of Ag-NHC and Au-NHC in A³-coupling reactions.

Run ^a	Catalyst	Aldehyde	Products	Yield ^b (%)
1	3a	p-formaldehyde	PAA1	25
2		cyclohexanecarboxaldehyde	PAA2	47
3		benzaldehyde	PAA3	23
4	4a	p-formaldehyde	PAA1	65
5		cyclohexanecarboxaldehyde	PAA2	52
6		benzaldehyde	PAA3	36
7	3b	p-formaldehyde	PAA1	86
8		cyclohexanecarboxaldehyde	PAA2	65
9		benzaldehyde	PAA3	60
10	4b	p-formaldehyde	PAA1	99
11		cyclohexanecarboxaldehyde	PAA2	99
12		benzaldehyde	PAA3	60

^a Reaction conditions: aldehyde (1.0 mmol), piperidine (1.2 mmol), phenylacetylene (1.5 mmol), (NHC)₂-M catalyst (3 mol %), 80 °C, nitrogen atmosphere, 6 h. ^b Conversions were determined by ¹H-NMR analysis (internal standard: 2-bromo mesitylene).

The reactions were carried out without the use of solvents, and the results were very interesting also from the point of view of sustainable chemistry. The yields were determined using ¹H-NMR analysis, by integration of the singlet signal of internal standard (2-bromomesitylene) at 6.89 ppm in CD₂Cl₂, with the signal of propargylic product (δ 3.43 ppm for N-(3-phenyl-2-propynyl) piperidine, PAA1, δ 3.11 ppm for N-(1-cyclohexyl-3-phenyl-2-propynyl) piperidine, PAA2, δ 4.79 ppm for PAA3 N-(1,3-diphenyl-2-propynyl) piperidine). As shown in Table 1, all complexes are capable of catalyzing the coupling of aldehyde, piperidine, and phenylacetylene. Silver complexes are less active than analog gold complexes (compare runs 1–3 with 7–9 and 4–6 with 10–12). Complexes **3a** and **4a**, which have N-heterocyclic carbene with hydrogens on the backbone, are less active than silver and gold complexes **3b** and **4b** with chlorine atoms on the backbone (compare runs 1–6 with 7–12). The greatest activity of complexes bearing a NHC ligand with two chlorine atoms on the backbone was also found in our previous work [37].

It was attributable to a major positive charge on the metal center, which makes it more electrophilic and more active to coordinate the phenylacetylene. Moreover, benzaldehyde is less reactive with respect to aliphatic aldehydes.

3. Biological Activities

3.1. Antibacterial Activity

The antibacterial activity of compounds containing Ag (**3a** and **4a**) and Au (**3b** and **4b**) were assessed by determining the MIC (minimal inhibitory concentration) according to the CLSI (Clinical and Laboratory Standards Institute) guidelines [48,49]. The compounds

were tested against both Gram positive *Staphylococcus aureus* and Gram-negative bacteria *Escherichia coli*. Another analog molecule, **1a**, previously demonstrated effective on *E. coli* [40] and pro-ligands **S1**, **S2**, **S3**, **S4** were also introduced as experimental controls [40]. We have also tested the analog gold complex **1b** and, furthermore, the **2a** and **2b**, which have chlorine on the backbone of the imidazole ring instead of hydrogen. Results are shown in Table 2. Compounds containing Ag were very effective on *E. coli*, with MIC values between 10 and 15 µg/mL, whereas, except for **3a** (MIC 50 µg/mL), up to the maximum concentration tested (150 µg/mL) they had no effect on the growth of *S. aureus*. As expected, **1a** exhibited high antimicrobial action against the tested strain of *E. coli* (MIC 6.5 µg/mL). On the contrary, all molecules containing Au were very efficient against the Gram-positive *S. aureus*, with very low MIC values. In particular, **2b** showed a minimum inhibition dose of 0.5 µg/mL. Unlike the compounds with Ag, the Au complexes were active towards both bacteria species. In fact, except for **1b** (MIC >15 µg/mL), all showed a fair antimicrobial activity towards Gram-negative (MIC 75–100 µg/mL), indicating for these latter properties of broader spectrum antimicrobials. For both bacterial genera, the pro-ligands **S1–S4** showed no growth inhibition effect up to the tested concentration of 200 µg/mL.

Table 2. Antimicrobial activity tested on representative Gram negative and positive bacteria.

Complex	MIC (Minimal Inhibitory Concentration) µg/mL (µM) ^a	
	<i>E. Coli</i> (Gram Negative)	<i>S. Aureus</i> (Gram Positive)
1a ^b	6.5 (14.9)	>150 (>343.2)
2a	10 (19.8)	>150 (>296.4)
3a	15 (32.1)	50 (107.0)
4a	15 (27.9)	>150 (>279.8)
1b	>150 (>345.1)	40 (92.0)
2b	75 (148.9)	0.5 (0.99)
3b	100 (215.2)	25 (53.8)
4b	75 (140.7)	2.5 (4.7)

Reported values were determined by three independent assays, each in triplicate. ^a 150 µg/mL is the highest concentration tested for each compound. ^b Complex **1a** was introduced as experimental control active versus *E. coli* strains.

3.2. Anticancer Activity

The complex **1a** was characterized as in rif. [40]. The Ag complexes **3a** and **4a** and Au complexes **3b** and **4b** were evaluated for their anticancer activity towards two human breast cancer cell lines, namely MCF-7 and MDA-MB-231 cells, and the human cervix carcinoma HeLa cells. As shown in Table 3, where the IC₅₀ values are listed, the silver complexes were more active compared to the gold ones. In particular, the Ag complexes **3a** and **4a** exerted the best anticancer activity towards the HeLa cells, possessing a comparable activity with IC₅₀ values of 12.2 ± 1.0 µM for **3a** and 11.9 ± 0.4 µM for **4a**. These compounds also exhibited good anticancer activity on the MCF-7 cells, but to a lower extent when compared to that displayed on HeLa cells, with IC₅₀ values of 20.3 ± 1.1 µM and 19.5 ± 0.9 µM, respectively. In contrast, Au complexes showed no anticancer activity in all the cancer cell lines tested, at least until the concentration reached 200 µM, except for Au complex **4b** that exhibited a good anticancer activity towards the MCF-7 cells (IC₅₀ = 12.2 ± 1.2 µM) and a very low activity on the triple negative MDA-MB-231 cells (IC₅₀ = 49.5 ± 0.7 µM). None of the gold-based complexes (**3b** and **4b**) were found to be active against HeLa. Moreover, we verified whether the pro-ligands by themselves could exert a cytotoxic activity on the same cancer cells. As a result, neither **S3** nor **S4** exerted anticancer effects, at least at doses equal to or below 200 µM. Once established that some of the new synthesized complexes possess interesting antibacterial and antitumor activities, we tested all the compounds on two human normal cells, the breast epithelial MCF-10A cells, and the embryonic kidney cells Hek-293, in order to determine their cytotoxic effect. The obtained results showed that none of the Au and Ag complexes affected the viability of the used normal cells up to

200 μM . The silver complexes were more active than Cisplatin, a well-known anticancer drug used as reference molecule [39]. Indeed, Cisplatin was able to reduce MCF-7 and HeLa cells viability with IC_{50} values of $36.2 \pm 1.0 \mu\text{M}$ and $16.2 \pm 1.1 \mu\text{M}$, respectively. Moreover, each metal complex did not affect the viability of the two normal cell lines, in contrast to Cisplatin, which showed a strong cytotoxicity ($\text{IC}_{50} = 80.7 \pm 1.0 \mu\text{M}$ on MCF-10A and $16.1 \pm 0.9 \mu\text{M}$ on Hek-293).

Table 3. IC_{50} values of the studied metal complexes (3a,b and 4a,b) and the ligands (S3 and S4) expressed in μM .

Sample	IC_{50} (μM)				
	MDA-MB-231	MCF-7	HeLa	MCF-10A	Hek-293
S3	>200	>200	>200	>200	>200
S4	>200	>200	>200	>200	>200
3a	>200	20.3 ± 1.1	12.2 ± 1.0	>200	>200
3b	>200	>200	>200	>200	>200
4a	>200	19.5 ± 0.9	11.9 ± 0.4	>200	>200
4b	49.5 ± 0.7	12.2 ± 1.2	>200	>200	>200
Cisplatin	28.9 ± 0.7	36.2 ± 1.0	16.2 ± 1.1	80.7 ± 1.0	16.1 ± 0.9

4. Experimental Design

4.1. Materials and Methods

All synthesis, manipulations, and characterization experiments were carried out under an oxygen and moisture free atmosphere using Schlenk techniques. Reagents were purchased from Sigma Aldrich and TCI Chemicals and used without any other operation of purification. Solvents were degassed and dehydrated under a nitrogen atmosphere by heating at reflux over suitable drying agents. NMR solvents (Euriso-Top products) were kept out of the light over molecular sieves. NMR spectra were recorded on Bruker AM 300 spectrometers (300 MHz for ^1H ; 75 MHz for ^{13}C) and a Bruker AVANCE 400 spectrometer (400 MHz for ^1H ; 100 MHz for ^{13}C). The samples were prepared by dissolving 20 mg of compounds in 0.6 mL of deuterated solvent. The chemical shifts of ^1H -NMR and ^{13}C -NMR spectra are referenced to tetramethylsilane (SiMe_4 , $\delta = 0$ ppm). The spectrum multiplicities are abbreviated in this manner: singlet (s), doublet (d), triplet (t), multiplet (m), and broad (br).

The elementary analyses for C, H, and N were performed according to standard microanalytical procedures, recording them by means of a Thermo-Finnigan Flash EA 1112.

The determination of the quantities of chloride and iodide was performed indirectly by reaction of AgNO_3 with the halogen and precipitation of the silver halide (AgX). It was dissolved with $\text{Na}_2\text{S}_2\text{O}_3$, and the silver content in the solution was determined by atomic flame absorption spectroscopy (FAAS), achieving the halogen content from the amount of silver. The mass of the organic compounds was determined by ESI-MS with a Waters Quattro Micro triple quadrupole mass spectrometer furnished with an electrospray ion source.

ESI-FT-ICR measurements of the complexes were carried out using a Bruker Solaris XR instrument. MALDI-MS mass spectra were recorded using a Bruker SolariX (Bruker Daltonik GmbH, Bremen, Germany) Fourier transform ion cyclotron resonance mass spectrometer equipped with a refrigerated 7 T actively shielded superconducting magnet (Bruker Biospin, Wissembourg, France). The samples were positively ionized using the MALDI ion source (Bruker Daltonik GmbH, Bremen, Germany). The laser power was 28%, and 22 laser shots were used for each scan. The mass range was set to m/z 200–3000. The mass spectra were calibrated externally, applying linear calibration and using a mix of peptide clusters in MALDI positive ionization mode. To improve the accuracy of the measurement, the spectra of the samples were recalibrated internally by matrix ionization (2,5-dihydroxybenzoic acid).

4.2. Synthesis

4.2.1. General Procedure for Synthesis of N-Heterocyclic Carbene Proligands (**S3** and **S4**)

Imidazolium salts **S3** and **S4** were synthesized following the synthetic strategy by Arnold and coworkers [41,42] and applying the procedures reported in literature by us [37–40,45,46]. Imidazole or 4,5-dichloro imidazole (1.0 eq) was reacted with phenylethylene oxide (1.2 eq) for 12 h at refluxing temperature, and subsequently 2-iodoethanol (2.0 eq) was added. The imidazolium salts were obtained by precipitation and washed with hexane (3 × 20 mL) and diethyl ether (3 × 30 mL).

4.2.2. Synthesis of Iodo [N-(2-Hydroxyethyl)-N'-(2-Hydroxy-2-Phenyl) Ethyl-Imidazole-2-Ylidene] (**S3**)

Imidazole (1.00 g, 15.4 mmol) and phenylethylene oxide (2.40 g, 18.5 mmol) were dissolved in 25 mL of CH₃CN and stirred at refluxing temperature for 12 h. Subsequently, the reaction mixture was cooled, and 2-iodoethanol (5.29 g, 30.8 mmol) was added. The mixture was warmed up to refluxing temperature and stirred for 8 h more. The product was recovered by removing the solvent and precipitation in cold acetone. The resulting powder (5.73 g, 15.9 mmol, yield 86%) was washed with hexane (3 × 20 mL) and diethyl ether (2 × 30 mL).

¹H-NMR (δ ppm, DMSO-d₆, 400 MHz): 9.12 (s, NCHN, 1H), 7.75–7.35 (m, aromatic hydrogens, 7H), 5.99 (d, OH, 1H), 5.17 (b, OH, 1H), 4.97 (t, CHOH, 1H), 4.46–4.25 (m, NCH₂CH₂OH, NCH₂CHOH, 4H), 3.75 (d, NCH₂CH₂OH, 2H). (in Supplementary Materials Figure S1)

¹³C-NMR (δ ppm, DMSO-d₆, 100 MHz): 141.28 (*ipso* carbon of aromatic ring), 136.75 (NCHN), 128.34, 127.82, 125.99 (aromatic carbons), 122.95, 122.28 (backbone carbons), 70.70 (CHOH), 59.36 (CH₂OH), 55.60 (NCH₂CHOH), 51.56 (NCH₂CH₂OH). (in Supplementary Materials Figure S2)

MALDI, (m/z): 233.12898 Dalton attributable to [C₁₃H₁₇N₂O₂]⁺. (in Supplementary Materials Figure S3)

4.2.3. Synthesis of Iodo [4,5-Dichloro N-(2-Hydroxyethyl) -N'-(2-Hydroxy-2-Phenyl) Ethyl-Imidazole-2-Ylidene] (**S4**)

Both 4,5-dichloro imidazole (1.00 g, 7.33 mmol) and phenylethylene oxide (1.32 g, 11.0 mmol) were dissolved in 25 mL of CH₃CN and stirred at refluxing temperature for 12 h. Subsequently, the reaction mixture was cooled, and 2-iodoethanol (2.52 g, 14.6 mmol) was added. The reaction mixture was warmed up to refluxing temperature and stirred for 8 h more. The product was recovered, removing the solvent and precipitation in cold acetone. The resulting powder (1.88 g, 4.39 mmol, yield 60%) was washed with hexane (3 × 20 mL) and diethyl ether (2 × 30 mL).

¹H-NMR (δ ppm, DMSO-d₆, 400 MHz): 9.51 (s, NCHN, 1H), 7.40–7.33 (m, aromatic hydrogens, 5H), 6.05 (d, OH, 1H), 5.18 (t, OH, 1H), 4.99 (t, CHOH, 1H), 4.49–4.34 (m, NCH₂CHOH, NCH₂CH₂OH; 4H), 3.77 (d, NCH₂CH₂OH, 2H) (in Supplementary Materials Figure S4).

¹³C-NMR (δ ppm, DMSO-d₆, 100 MHz): 140.44 (*ipso* carbon of aromatic ring), 137.39 (NCHN), 128.47, 128.10, 125.96 (aromatic carbons), 118.91, 118.47 (backbone carbons), 69.80 (CHOH), 58.18 (CH₂OH), 54.85 (NCH₂CHOH), 51.08 (NCH₂CH₂OH). (in Supplementary Materials Figure S5)

MALDI: (m/z): 301.05142 Dalton attributable to [C₁₃H₁₅Cl₂N₂O₂]⁺ (in Supplementary Materials Figure S6).

4.2.4. General Procedure for the Synthesis of Silver(I) N-Heterocyclic Carbene Complexes (**3a** and **4a**)

The imidazolium salt **S3** or **S4** (1 eq) was suspended in *dry*-dichloromethane (25 mL) and molecular sieves 4 Å. Ag₂O (0.7 eq) was added to mixture, and it was stirred for 4 h at room temperature with no light. It was then filtered on a pad of Celite to eliminate the

silver iodide byproduct and the molecular sieves. The complexes (**3a** and **4a**) were obtained after the removal of the solvent in vacuo.

4.2.5. Synthesis of Iodo

[N-(2-Hydroxyethyl)-N'-(2-Hydroxy-2-Phenyl)Ethyl-Imidazole-2-Yliden] Silver(I) (**3a**)

The silver complex **3a** was obtained by reaction of imidazolium salt **S3** (0.508 g; 1.41 mmol) and Ag₂O (0.202 g; 0.914 mmol) in dry dichloromethane (25 mL) in the presence of 4 Å molecular sieves, at room temperature and in dark conditions for 4 h. The reaction mixture was filtered by celite and solvent was evaporated in vacuo. The residual powder was washed by dry diethyl ether (3 × 30 mL), and dried by reduced pressure to lead the silver complexes as a white powder (0.330 g, 0.71 mmol, yield: 50%).

¹H-NMR (δ ppm, DMSO-d₆, 400 MHz): 7.38–7.31 (m, aromatic hydrogens, 7H), 5.88 (b, CHO, 1H) 5.07 (b, CH₂OH, 1H), 4.99 (b, NCH₂CHO, 1H), 4.33–4.27 (m, NCH₂CHO, NCH₂CH₂OH, 4H), 3.73 (b, NCH₂CH₂OH, 2H) (in Supplementary Materials Figure S7).

¹³C-NMR (δ ppm, DMSO-d₆, 100 MHz): 181.23–179.28 (dd, ¹J_{C-107-Ag} = 182.3 Hz, ¹J_{C-109-Ag} = 209.1 Hz), 142.53 (*ipso* carbon aromatic ring), 128.10, 127.10, 126.02 (aromatic carbons), 122.40, 121.85 (backbone carbons), 72.46 (CHO), 61.19 (CH₂OH), 58.40 (NCH₂CHO), 53.62 (NCH₂CH₂OH) (in Supplementary Materials Figure S8).

ESI-MS (CH₃CN, m/z): 573.14690 Da attributable to [C₂₆H₃₂AgN₄O₄]⁺ (in Supplementary Materials Figure S9).

4.2.6. Synthesis of Iodo [4,5-Dichloro N-(2-Hydroxyethyl) -N'-(2-Hydroxy-2-Phenyl) Ethyl-Imidazole-2-Yliden] Silver(I) (**4a**)

The silver complex **4a** was synthesized by reaction of iodo [4,5-dichloro N-(2-hydroxyethyl) -N'-(2-hydroxy-2-phenyl) ethyl-imidazole-2-ylidene] (0.530 g; 1.23 mmol) with silver oxide (0.171 g; 0.74 mmol) in dry dichloromethane (25 mL) in the presence of 4 Å molecular sieves at room temperature for 4 h in the dark. The reaction mixture was then filtered on a pad of Celite, the solvent was removed, and the residual powder was washed with dry diethyl ether (3 × 30 mL) to give a light-yellow powder (0.362 g, 0.677 mmol, yield: 55%).

¹H-NMR (δ ppm, DMSO-d₆, 400 MHz): 7.38–7.28 (m, aromatic hydrogens, 5H), 5.89 (s, CHO, 1H), 5.07 (b, CH₂OH, 1H) 4.99 (b, NCH₂CHO, 1H), 4.33–4.27 (m, NCH₂CHO, NCH₂CH₂OH, 4H), 3.74 (b, NCH₂, 2H) (in Supplementary Materials Figure S10).

¹³C-NMR (δ ppm, DMSO-d₆, 100 MHz): 183.67 (C_{carbene}-Ag), 141.51 (*ipso* carbon aromatic ring), 128.31, 127.72, 126.08 (aromatic carbons), 117.10, 116.74 (backbone carbons), 71.63 (CHO), 60.28 (CH₂OH), 56.99 (NCH₂CHO), 52.57 (NCH₂CH₂OH) (in Supplementary Materials Figure S11).

ESI-MS (CH₃CN, m/z): 707.99716 Dalton attributable to [C₂₆H₂₈AgCl₄N₄O₄]⁺.

4.2.7. General Procedure for the Synthesis of Gold(I) N-Heterocyclic Carbene Complexes (**3b** and **4b**)

Following the literature, gold complexes **3b** and **4b** were prepared by *trans*-metalation route [2–10]. The imidazolium salt (1.0 eq) was reacted with the silver oxide (0.7 eq). The mixture was then filtered and solution was added (chloro(dimethylsulfide)gold (I)) to give gold NHC complexes.

4.2.8. Synthesis of Chloro [N-(2-Hydroxyethyl)-N'-(2-Hydroxy-2-Phenyl)Ethyl-Imidazole-2-Yliden] Gold(I) (**3b**)

The imidazolium salt **S3** (0.500 g; 1.40 mmol) and Ag₂O (0.202 g; 0.98 mmol) were dissolved in dry dichloromethane (25 mL) containing 4 Å molecular sieves. The mixture was stirred at room temperature with exclusion of light for 4 h. It was then filtered by a pad of Celite to remove the silver iodide byproduct and molecular sieves. Chloro(dimethylsulfide)gold(I) (0.415 g, 1.40 mmol) was added to solution and the reaction mixture was stirred for other 4 h, at room temperature under dark conditions. Afterward, the mixture was further filtered to eliminate AgI byproduct. The solution was dried a

reduced pressure to lead a yellow powder. The residual was washed with diethyl ether obtaining the gold complex **3b** (0.260 g, 0.56 mmol, yield: 40%).

$^1\text{H-NMR}$ (δ ppm, DMSO- d_6 , 400 MHz): 7.40–7.36 (m, aromatic hydrogens, 7H), 5.81 (m; CHOH, 1H) 5.08–5.02 (m, NCH₂CH₂OH, NCH₂CHOH, 4H), 3.74 (d, NCH₂CH₂, 2H) (in Supplementary Materials Figure S12).

$^{13}\text{C-NMR}$ (δ ppm, DMSO- d_6 , 100 MHz): 168.37 ($C_{\text{carbene-Au}}$), 142.07 (*ipso* carbon aromatic ring), 128.30, 127.79, 127.61 (aromatic carbons), 122.33, 121.56 (backbone carbons), 72.36 (CHOH), 60.44 (CH₂OH), 57.79 (NCH₂CHOH), 53.00 (NCH₂CH₂OH) (in Supplementary Materials Figure S13).

ESI-MS (CH₃CN, *m/z*): 661.20838 Dalton attributable to [C₂₆H₃₂AuN₄O₄]⁺ (in Supplementary Materials Figure S14).

4.2.9. Synthesis of Chloro [4,5-Dichloro (N-(2-Hydroxyethyl)-N'-(2-Hydroxy-2-Phenyl) Ethyl-Imidazole-2-Yliden] Gold(I) (**4b**)

At suspension of imidazolium salt S4 (0.530 g; 1.23 mmol) in dry dichloromethane (25 mL) silver oxide (0.198 g; 0.861 mmol) was added, and the mixture was stirred for 4 h in dark conditions to give the silver (I) complex. The mixture was filtered to remove the AgI, and the gold precursor (0.362 g; 1.23 mmol) was added to grey solution. The mixture was stirred for other 4 h in the dark. It was then filtered, and the yellow solution was dried in vacuo, and the gold complex **4b** was obtained after washing with diethyl ether (yield: 61%, corresponding to 0.400 g, 0.75 mmol).

$^1\text{H-NMR}$ (δ ppm, DMSO- d_6 , 400 MHz): 7.38–7.28 (m, 5H aromatic hydrogens), 5.89 (s, CHOH, 1H), 5.07–4.99 (m; CH₂OH; 2H) 4.33–4.28 (m, NCH₂CH₂OH, 2H), 3.74 (d, NCH₂, 2H) (in Supplementary Materials Figure S15).

$^{13}\text{C-NMR}$ (δ ppm, DMSO- d_6 , 100 MHz): 170.90 ($C_{\text{carbene-Au}}$) 141.35 (*ipso* carbon aromatic ring), 128.4, 127.94, 125.08 (aromatic carbons), 117.20, 116.74 (backbone carbons), 72.14 (CHOH), 59.85 (CH₂OH), 56.89 (NCH₂CHOH), 52.37 (NCH₂CH₂OH) (in Supplementary Materials Figure S16).

ESI-MS (CH₃CN, *m/z*): 709.05720 Dalton attributable to [C₂₆H₂₈AuCl₄N₄O₄]⁺ (in Supplementary Materials Figure S17).

4.3. General Procedure for A³ Coupling (Aldehyde, Amine, Alkyne) Reaction: General Procedure of A³ Coupling Reaction Promoted by M-NHC Catalysts

In a 10 mL Schlenk tube, the suitable aldehyde (1.0 mmol), piperidine (1.2 mmol), and phenylacetylene (1.5 mmol) were introduced with 2-bromomesitylene as internal standard (1.0 mmol) and the catalyst (3% mol). The reaction mixture was stirred for 6 h at 80 °C. It was then cooled at room temperature, and CH₂Cl₂ was added. The organic phase was dried on magnesium sulfate and filtered. The catalytic activity was evaluated by $^1\text{H-NMR}$, integrating the signal at 6.89 (2H of internal standard) and protons in α to nitrogen atom of propargylic amine, i.e., at δ 3.43, 2H for *N*-(3-phenyl-2-propynyl) piperidine PAA1; at δ 3.11, 1H for 1-(1-cyclohexyl-3-phenyl-2-propynyl) piperidine PAA2; at δ 4.79, 1H for *N*-(1,3-diphenyl-2-propynyl) piperidine PAA3, respectively. The NMR characterization of coupling products is reported in ref. [41].

4.4. Antibacterial Activity

Microbiological Assays and Bacterial Strains

The antimicrobial activity of compounds containing both Ag and Au (**1a–4a** and **1b–4b**) was estimated by determining the minimal growth inhibitory concentrations (MIC), according to the indications given in the guidelines of the Clinical and Laboratory Standards Institute (CLSI) [17,22]. All molecules were tested in the concentration range of 0–150 $\mu\text{g/mL}$ (0, 0.5, 1, 2, 3, 4.5, 6.5, 7.5, 10, 15, 25, 40, 50, 75, 100, 150). Antibacterial compound **1a** [40] and pro-ligands **S1**, **S2**, **S3**, **S4** were introduced as experimental controls. Briefly, the bacteria were suspended in Luria-Bertani (LB) broth (tryptone, 10 g/L; yeast extract 10 g/L; sodium chloride, 5 g/L, pH 7) at a density of $5 \cdot 10^5$ CFU/mL and incubated

in the presence of the different concentrations of each compound at 37 °C, with constant shaking (250 rpm). After 24 h the effects on growth were evaluated by turbidity, measuring the optical density (OD) at 600 nm. The MIC was defined as the lowest concentration that did not change the turbidity of the sample with respect to time 0. For each compound, the MIC was repeated in three independent experiments, each in triplicate.

Bacteria used for microbiological assays were *Escherichia coli* (strain JM109) and the pathogenic *Staphylococcus aureus* (hospital isolate), representative of Gram negative and Gram positive, respectively. *E. coli* was purchased from Promega (<http://www.promega.com/products>, accessed on 28 October 2021). The *S. aureus* strain was obtained from the microbial collection deposited in the microbiology laboratory of the University of Salerno directed by Prof. G. Vigliotta.

4.5. Anticancer Activity

4.5.1. Cell Culture

The cell lines employed in this work (MCF-7, MDA-MB-231, HeLa, MCF-10A, and Hek-293) were purchased from American Type Culture Collection (ATCC, Manassas, VA, USA). MCF-7 and MDA-MB-231 human breast cancer cells were maintained in Dulbecco's modified Eagle's medium/nutrient mixture Ham F-12 (DMEM/F12), supplemented with 5% fetal bovine serum (FBS, Thermo Fisher Scientific, Milan, Italy) and 1% penicillin/streptomycin. HeLa human epithelial cervix carcinoma cells were cultured in minimum essential Eagle's Medium (MEM), supplemented with 10% FBS, 1% L-glutamine, 1% penicillin/streptomycin, and 1% nonessential amino acids (NEAA). MCF-10A human mammary epithelial cells were cultured in DMEM/F12 medium, supplemented with 5% horse serum (HS, Thermo Fisher Scientific, Milan, Italy), 1% penicillin/streptomycin, 0.5 mg mL⁻¹ hydrocortisone, 20 ng mL⁻¹ human epidermal growth factor (hEGF), 10 mg mL⁻¹ insulin, and 0.1 mg mL⁻¹ cholera enterotoxin (Sigma-Aldrich, Milan, Italy). Hek-293 human embryonic kidney cells were cultured in DMEM, supplemented with 10% FBS, 1% L-glutamine, and 1% penicillin/streptomycin. Cells were maintained at 37 °C with 5% CO₂ and periodically screened for contamination.

4.5.2. MTT Assay

The in vitro anticancer activity of all of the studied compounds were evaluated using the MTT assay as already described [40]. The treatment was carried out exposing the cells to the target compounds dissolved in DMSO at seven concentrations (0.1, 1, 5, 10, 50, 100, and 200 µM) for 72 h. MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide) was added for 2 h at 37 °C (final concentration 0.5 mg/mL). The formazan crystals were then dissolved in DMSO, and the optical density was measured at 570 nm using a microplate reader. All the calculations were performed in triplicate, and the results were represented as the percent (%) of basal. The IC₅₀ values were calculated using curve fitting GraphPad Prism 9 software (GraphPad Software, La Jolla, CA, USA) with nonlinear regression. The values represent the mean ± standard deviation (n = 3).

5. Conclusions

In summary, we prepared silver and gold complexes with NHC ligands of general formula [M(NHC)₂]⁺ [MX₂]⁻. The complexes were used as catalysts for the A³-coupling reactions of phenylacetylene, piperidine, and three different aldehydes (p-formaldehyde, cyclohexanecarboxaldehyde, and benzaldehyde) to afford propargylamines. They showed moderate to high activity. Better performances were recorded with Au(I) complex **4b** in the reaction of phenylacetylene and piperidine with p-formaldehyde or cyclohexanecarboxaldehyde.

The antimicrobial activity of complexes **3a,b** and **4a,b** was evaluated against *Escherichia coli* and *Staphylococcus aureus* as representatives of Gram-negative and Gram-positive bacteria and compared with that of complexes **1a,b** and **2a,b**. The most active compounds against *E. coli* were the Ag(I) complexes (MIC values range from 6.5–15 µg/mL), whereas against *S. aureus* were the Au(I) complexes (MIC values range from 0.5 to 40 µg/mL).

Cytotoxicity of the pro-ligands (**S3** and **S4**) and **3a,b** and **4a,b** complexes were studied by means of a colorimetric assay (MTT assay) towards two human breast cancer cell lines, namely MCF-7 and MDA-MB-231, and human cervical carcinoma HeLa cells. The lowest inhibitory concentration values were verified for **4a** (HeLa cells, $IC_{50} = 11.9 \pm 0.4 \mu\text{M}$) and **4b** (MCF7, $IC_{50} = 12.2 \pm 1.2 \mu\text{M}$).

It is noteworthy to mention that the complexes are highly selective towards cancer cells and bacterial cells. In fact, treatments up to 72 h with the synthesized complexes do not affect the viability of the normal MCF-10A breast epithelial cells and the embryonic kidney cells Hek-293; moreover, at non-toxic doses for these cells they have high antimicrobial activity.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/catal12010018/s1>. Figure S1: $^1\text{H-NMR}$ of **S3**; Figure S2: $^{13}\text{C-NMR}$ of **S3**; Figure S3: ESI-MS of **S3**; Figure S4: $^1\text{H-NMR}$ of **S4**; Figure S5: $^{13}\text{C-NMR}$ of **S4**; Figure S6: MALDI-MS of **S4**; Figure S7: $^1\text{H-NMR}$ of **3a**; Figure S8: $^{13}\text{C-NMR}$ of **3a**; Figure S9: ESI-MS of **3a**; Figure S10: $^1\text{H-NMR}$ of **4a**; Figure S11: $^{13}\text{C-NMR}$ of **4a**; Figure S12: $^1\text{H-NMR}$ of **3b**; Figure S13: $^{13}\text{C-NMR}$ of **3b**; Figure S14: MALDI OF **3b**; Figure S15: $^1\text{H-NMR}$ of **4b**; Figure S16: $^{13}\text{C-NMR}$ of **4b**; Figure S17: MALDI of **4b**.

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