



Novel sterically hindered cannabinoid CB₁ receptor ligands

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abstract

In the present study, 11 novel N-(3,3-diphenyl)propyl-2,2-diphenylacetamide derivatives (4a–d and 9a–g) and six triphenylacetamides (10a–c and 11a–c) were synthesised and tested as ligands of cannabinoid CB₁ and CB₂ receptors. All compounds exhibited affinity for CB₁ and CB₂ receptors. Four compounds (4b, 9a, 9b, and 11a) showed selectivity for CB₁ versus CB₂ receptors, although only the N-(3,3-diphenyl)propyl-2,2-diphenylacetamide (4b) can be considered a potent CB₁ ligand (K_i 58 nM). It was 140-fold selective over CB₂ receptors (K_i 7800 nM) and behaved as an inverse agonist by stimulating forskolin-induced cAMP formation in mouse N18T2 neuroblastoma cells. This compound is the rst of a novel class of tetraphenyl CB₁ ligands that, in view of its easy synthesis and high affinity for CB₁ receptors and despite its sterical hindrance, will be useful for the design of new blockers of this therapeutically exploitable receptor type.

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1. Introduction

Mammalian tissues contain at least two types of cannabinoid receptors, CB₁ and CB₂,^{1,2} that are coupled to G-proteins of G_{i/o} type. CB₁ receptors, cloned in 1992, are mostly expressed in the central nervous system, but also in peripheral tissues including immune cells, the reproductive system, the gastrointestinal tract and the lung, while CB₂ receptors, cloned in 1993, are most abundant in the immune system, that is, in tonsils, spleen, macrophages, and lymphocytes (B-cells and natural killer cells).^{3–5} It has been widely shown that there are several pathophysiological conditions, including pain,⁶ inflammation,⁷ liver diseases,⁸ and obesity,⁹ in which blocking the cannabinoid receptors might be beneficial. In fact, many CB₁ receptor antagonists have been developed so far^{10,11} and some of them are in clinical trials for the treatment of several disorders. One of these drugs, the CB₁ receptor antagonist/inverse agonist rimonabant (SR141716A) (Fig. 1) (1) belongs to the class of diarylpyrrole antagonists, including also other widely used pharmacological tools, such as AM-251 (2) and AM-281 (3). Rimonabant has been recently approved for marketing in the Eas as an adjunct to exercise and diet for the treatment of obesity and metabolic syndrome, and has proved useful to reduce body weight, low HD-cholesterol and high triglyceride levels, as well as

high glycemia, in obese patients, but also hallmarks of type-2 diabetes in treated and untreated patients.^{12–14} More recently, two other CB₁ receptor antagonists/inverse agonists have undergone clinical trials for the treatment of obesity: S319 (4), whose structure still resembles that of rimonabant, and M-0364 (5), which instead belongs to a different class of acyclic compounds and exhibits higher affinity at CB₁ receptors, and higher selectivity versus CB₂ receptors, than rimonabant.¹⁵

The chemical structures of these previously developed compounds (Fig. 1) show a striking difference from those of both D⁹-tetrahydrocannabinol (6) (Fig. 2), the Cannabis sativa natural component from which the cannabinoid receptors were discovered, and the endo cannabinoids anandamide (7) and 2-arachidonoyl-glycerol (8). These two naturally occurring classes of CB₁ receptor ligands, in fact, although containing pharmacophores found also in the various synthetic antagonists, are much less sterically hindered than rimonabant, S319, and M-0364.

On the basis of this background, we wondered if it would be possible to obtain new CB₁ receptor ligands with even higher sterical hindrance, and for this purpose we have synthesised eleven novel N-(3,3-diphenyl)propyl-2,2-diphenylacetamide derivatives (4a–d and 9a–g) and six triphenylacetamides (10a–c and 11a–c). Using the very simple synthetic procedure shown in Scheme 1, we obtained a-substituted acetamide derivatives 4a–d.

The synthesis of the 2-(4-substituted phenyl)-2-phenyl-N-(3,3-diphenylpropyl)acetamide 9a–g proceeds from a monosubstituted

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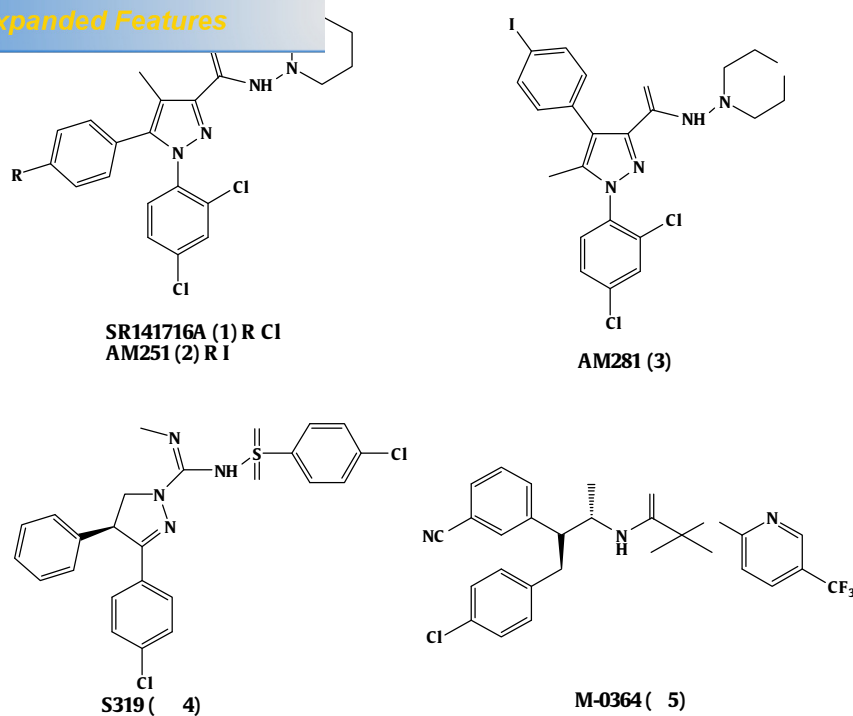


Figure 1. Chemical structures of some CB₁ receptor antagonists/inverse agonists.

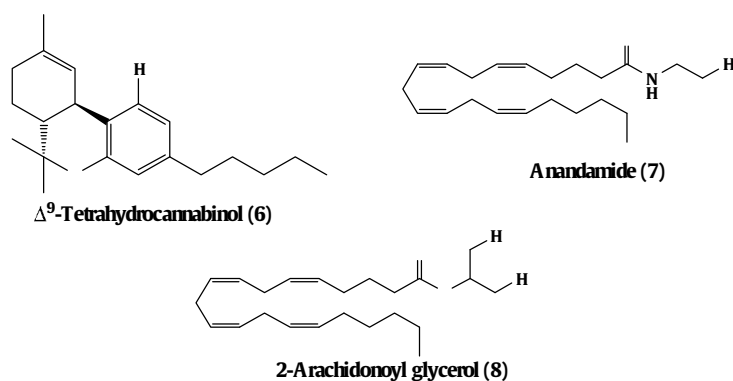
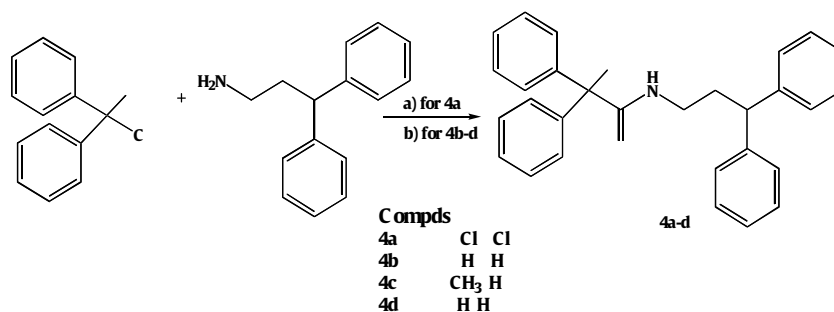


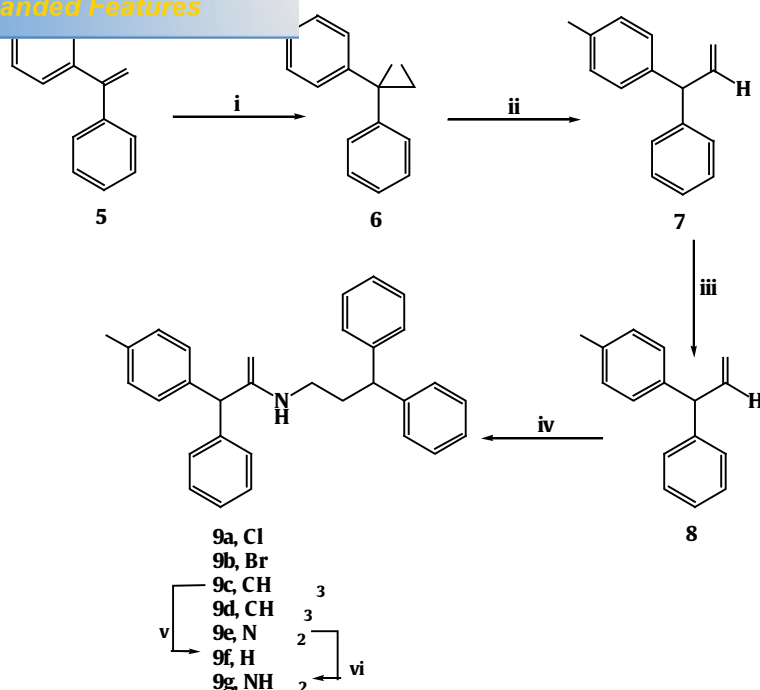
Figure 2. Chemical structures of D⁹-tetrahydrocannabinol and the endocannabinoids anandamide and 2-arachidonoyl glycerol.



Scheme 1. Schematic procedure for the one-step synthesis of 4a–d. Reagents and conditions (a) dry DCM, dry TEA, rt, 4 h (b) DCC, HBT, DCM, rt, 16 h.

diphenylacetic acid¹⁶ (Scheme 2). Homologation of commercially available 4-benophenones 5 (Cl, Br, CH₃, and N₂) by sodium hydride and trimethylsulfonium iodide gave the unstable epoxides 6, which were immediately converted into the aldehydes 7 by the action of BF₃ etherate.^{16,17} Then oxidation with Jones reagent¹⁸ converted the aldehydes 7 into the corresponding

acids 8 which, after amidation with 3,3-diphenylpropylamine, yielded the target compounds 9a–e (Scheme 2). The phenol derivative 9f was produced from the reaction of the methoxyphenyl derivative 9c with trimethylsilyl chloridesodium iodide¹⁹ (Scheme 2). Reduction of the nitro group of 9e by nHCl produced the amine derivative 9g²⁰ (Scheme 2).



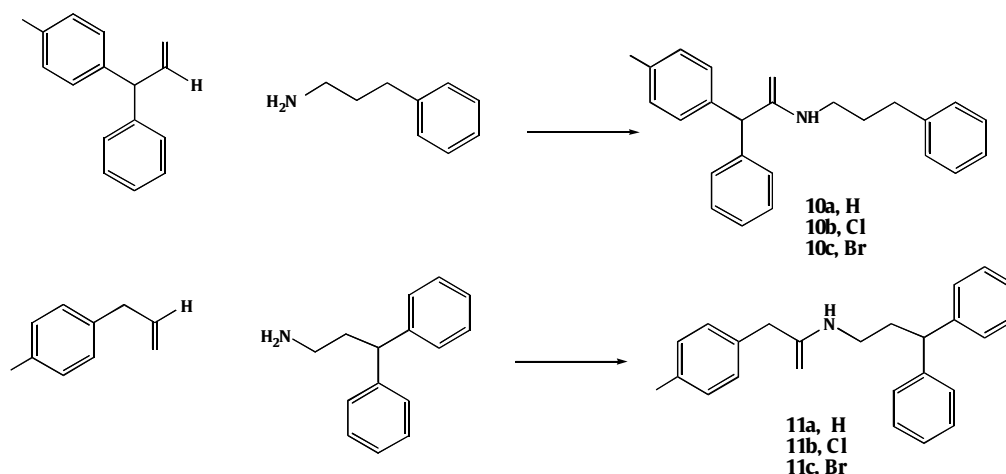
Scheme 2. Procedure for the multi-step synthesis of compounds 9a–g. Reagents and conditions (i) NaH, TMS-I, THF (ii) BF₃–Et₂, benene (iii) onesreagent, isopropyl alcohol, acetone (iv) 3,3-diphenylpropylamine, CDI, DMAP (v) NaI, TMS–Cl, CH₃CN (vi) n dust, HCl concd, EtH absolute.

Finally, we synthesised the six triphenylacetamides N-(3-phenyl)propyl-2,2-diphenylacetamide (10a), N-(3-phenyl)propyl-2-(4-chlorophenyl)-2-phenylacetamide (10b), N-(3-phenyl)propyl-2-(4-bromophenyl)-2-phenylacetamide (10c), N-(3,3-diphenyl)propyl-2-phenylacetamide (11a), N-(3,3-diphenyl)propyl-2-(4-chlorophenyl)acetamide (11b), and N-(3,3-diphenyl)propyl-2-(4-bromophenyl)acetamide (11c) by the following simple one-step synthetic procedure (Scheme 3).

The new compounds were tested for their affinities for human recombinant cannabinoid receptors CB₁ and CB₂, and the corresponding observed IC_{50} (nM) values are shown in Table 1. The functional activity of the most potent CB₁ ligand (4b) was also assessed at CB₁ receptors by studying its effect on forskolin-induced cAMP formation in mouse N18T2 neuroblastoma cells (Fig. 3). The most potent CB₁ cannabinoid receptor ligand in the present study was the N-(3,3-diphenyl)propyl-2,2-diphenylacetamide (4b), whose

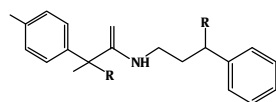
affinity for CB₁ receptors (IC_{50} 58 nM) was higher than that of anandamide (IC_{50} 89 nM). This compound also showed high (~140-fold) selectivity versus CB₂ receptors (IC_{50} 7900 nM). Compounds (4a, 4c–d) obtained from 4b, by introducing various substituents (Cl, CH₃, and H, respectively) in position 4 to the amide group, although showing still good affinity for CB₁, lost the selectivity versus CB₂ cannabinoid receptors.

The introduction of various substituents (Cl, Br, CH₃, CH₃, N₂, H, and NH₂) in para on one of the two aromatic rings closer to the amide functionality of the parent compound 4b led to more cannabinoid receptor ligands (9c–g), some of which (9a–b) were less selective (~16-fold) for CB₁ over CB₂ receptors. In addition, to test the effect of sterical hindrance on the binding to the cannabinoid receptor site, we eliminated first one of the two aromatic rings on the acyclic portion, and then one of those closer to the amide group of the selective compounds 4b, 9a, and 9b, thereby obtaining the six tri-



Scheme 3. Schematic procedure for the one-step synthesis of 10a–c and 11a–c. Reagents and conditions dry DCM, CDI, DMAP, rt, 4 h.

CB₂ receptors



4a-d, 9a-g, 10a-c and 11a-c

R	R	R	R	hCB ₁ (i, 1 M)	hCB ₂ (i, 1 M)	
4a	H	Cl	Phe	Phe	0.28 0.02	1.4 0.2
4b	H	H	Phe	Phe	0.058 0.01	7.9 0.3
4c	H	CH ₃	Phe	Phe	0.56 0.02	0.29 0.03
4d	H	H	Phe	Phe	2.2 0.15	2.3 0.2
9a	Cl	H	Phe	Phe	0.56 0.03	7.9
9b	Br	H	Phe	Phe	0.22 0.02	7.9
9c	CH ₃	H	Phe	Phe	0.56 0.04	0.79 0.04
9d	CH ₃	H	Phe	Phe	0.56 0.03	0.65 0.03
9e	N ₂	H	Phe	Phe	0.56 0.05	0.79 0.05
9f	H	H	Phe	Phe	0.56 0.02	1.1 0.2
9g	NH ₂	H	Phe	Phe	0.9 0.1	1.2 0.1
10a	H	H	H	Phe	0.56 0.05	7.9
	H	H	Phe	H	3.4 0.2	7.9
10b	Cl	H	Phe	H	2.2 0.2	2.4 0.2
10c	Br	H	Phe	H	1.9 0.1	1.8 0.2
11a	H	H	H	Phe	0.56 0.05	7.9
11b	Cl	H	H	Phe	1.6 0.1	2.4 0.3
11c	Br	H	H	Phe	0.8 0.02	1.3 0.1
AM251					0.0023 0.001	0.11 0.02
Rimonabant					0.008 0.001	0.79 0.1
SR144528					5.6	0.0054 0.001

Data represent mean values SEM for at least three separate experiments performed in duplicate and expressed as i , (1 M). AM251, CB₁ reference compound SR144528, CB₂ reference compound. Note Products commercially available. Registration numbers(4b) 339283-58-8, (10a) 353471-19-9, (11a) 543711-37-1, (11b) 560080-39-9, and (11c)749904-13-0.

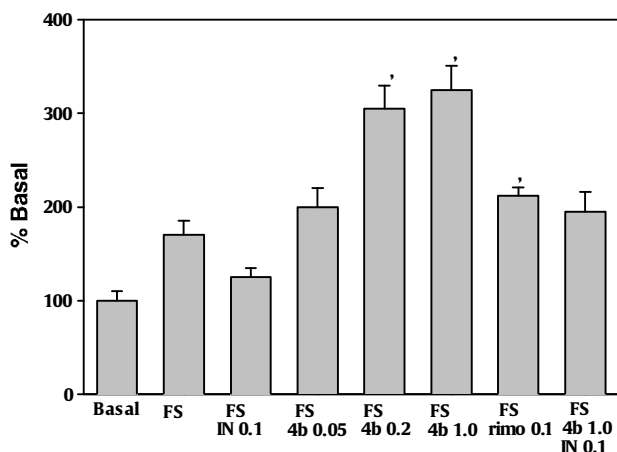


Figure 3. Effect of 4b (0.05, 0.2, and 1.0 μM) on forskolin (FS, 1 μM)-induced cAMP formation in intact N18T2 cells ($p < 0.05$ vs basal). The effects of IN55,212-2 (0.1 μM) and rimonabant (rimo, 0.1 μM) are shown as a comparison. * $p < 0.05$, 0.01 versus Basal. † $p < 0.05$ versus FS.

phenylacetamides 10a–c and 11a–c, respectively. This led to a significant reduction of the affinity and, with the exception of 10a and 11a, to the loss of the selectivity for the CB₁ receptors. It must be emphasized, however, that compounds 9a–g and 10b–c all contain an asymmetric center. Since we only determined the binding activity of the enantiomeric mixtures, the pure enantiomers might have exhibited different i values in the binding assays.

Finally, in order to establish the functional activity of the most potent and selective CB₁ ligand (4b), we tested its effect on forskolin-induced cAMP formation in intact N18T2 neuroblastoma cells, which constitutively and selectively express the CB₁ recep-

tor.²¹ As shown in Figure 3, the compound was found to stimulate cAMP formation in the presence of forskolin, as would be expected from an inverse agonist in this assay. However, the compound (0.05–1 μM) did not significantly elevate cAMP levels in the absence of forskolin (not shown). As expected, in the same assay, the CB₁/CB₂ agonist IN55,212-2 (0.1 μM) inhibited forskolin-stimulated cAMP formation, whereas the CB₁ inverse agonist, rimonabant (0.1 μM), produced a stimulation of forskolin effect. IN55,212-2 (0.1 μM) also blocked the effect of 4b (1 μM) on cAMP formation (Fig. 3), thus suggesting that the effect of 4b was mediated by CB₁ receptors.

The finding of 4b demonstrates that it is still possible to obtain high affinity and selective CB₁ receptor ligands by making compounds that are even more sterically hindered than rimonabant, S319, and M-0364. However, it is clear from our data that, although 4b maintains strong selectivity toward CB₂ receptors and functional activity as an antagonist/inverse agonist, it shows at least a 10-fold lower affinity toward the CB₁ receptor than these previously developed compounds, thus suggesting that the binding site of this receptor will probably not accept ligands with bigger hindrance. Interestingly, our new compounds resemble previously reported non rigid structures that were also shown to be CB₁ receptor inverse agonists.²²

In summary, we have described here the synthesis and pharmacological activity in vitro of a new class of sterically hindered CB₁ receptor ligands. The finding of 4b will be useful for future studies exploring further the structural requirements of the CB₁ receptor binding site. Furthermore, 4b will serve as a template for the development of new CB₁ inverse agonists, by capitalizing on the four phenyl groups present in this new molecule, which can be variedly derivatized as previously demonstrated by the several derivatives of rimonabant available to date.

2. Experimental

Melting points were taken on a allenkamp melting point apparatus and are uncorrected. ¹H NMR spectra were recorded with a Bruker Avance 300 MHz spectrometer. Mass spectra of compounds 4a–d, 9a–g, 10a–c, and 11a–c were obtained by C–MSMS analysis carried out via liquid chromatography–electrospray–ion trap–time of flight (C–ESI–IT–ToF) by using an IT–ToF mass spectrometer (Shimadzu) in conjunction with an C-20AB (Shimadzu). The ToF analyser allowed the determination of the molecular mass with high resolution. Chromatographic separations were performed on silica gel column (ieselgel 40, 0.040–0.063 mm, Merck). Reactions and product mixtures were routinely monitored by thin-layer chromatography (TC) on silica gel precoated F₂₅₄ Merck plates. All new compounds were ~98pure.

2.1. Synthesis of compounds 4a–d

2.1.1. N-(3,3-Diphenyl)propyl-2-chloro-2,2-diphenylacetamide (4a)
Dry triethylamine (130 mg, 1.28 mmol) was added to a stirred solution of 2-chloro-2,2-diphenylacetyl chloride (285 mg, 1.07 mmol), and 3,3-diphenylpropylamine (270 mg, 1.28 mmol) in dry dichloromethane (4 mL) at room temperature. After 4 h, the solvent was removed under reduced pressure, and the residue was taken up in EtAc and washed with brine. The organic portion was dried (Na₂S₄), solvent was evaporated and resulting residue was purified by silica gel column chromatography (n-hexane/EtAc, 7/3) to give the title compound (273 mg, 58%) as a white solid mp 125 °C ¹H NMR (CDCl₃) δ 2.38 (s, 2H, 7.5 H), 3.37 (s, 2H, 6.6 H), 3.95 (t, 1H, 7.9 H), 7.02 (br s, 1H), 7.23–7.46 (m, 20H). HR m/z 462.1595 correlates with the chemical formula C₂₉H₂₆NClNa within ≤1 ppm.

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N,N'-dicyclohexylcarbodiimide (500 mg, 2.42 mmol) was added to a stirred mixture of diphenylacetic acid (460 mg, 2.17 mmol), HBT (327 mg, 2.42 mmol), and 3,3-diphenylpropylamine (511 mg, 2.42 mmol) in dry dichloromethane (15 mL) at room temperature. After 16 h, the reaction mixture was filtered through a pad of Celite, and the filtrate was evaporated under reduced pressure. The resulting residue was taken up with EtAc (15 mL) and washed with 2 N NaH solution (2 × 15 mL), 2 N HCl solution (2 × 15 mL) and brine (2 × 15 mL), then the organic phase was dried (Na₂S₄), and evaporated under reduced pressure. The resulting residue was purified by silica gel column chromatography (n-hexane/EtAc, 7/3) to give the title compound (764 mg, 87%) as a white solid mp 130 °C ¹H NMR (CDCl₃) δ 2.28 (s, 2H, 7.2 H), 3.27 (s, 2H, 6.5 H), 3.86 (t, 1H, 7.8 H), 4.88 (s, 1H), 5.57 (br s, 1H), 7.19–7.38 (m, 20H). HR m/z 428.2007 correlates with the chemical formula C₂₉H₂₇NNa within ≤5 ppm.

2.1.3. N-(3,3-Diphenyl)propyl-2-methyl-2,2-diphenylacetamide (4c)

Starting from 2,2-diphenylpropionic acid (350 mg, 1.55 mmol), the title compound (530 mg, 82%) was obtained as reported for 4b as a white solid mp 133 °C ¹H NMR (CDCl₃) δ 1.99 (s, 3H), 2.25 (s, 2H, 7.4 H), 3.22 (s, 2H, 6.6 H), 3.77 (t, 1H, 7.9 H), 5.48 (br s, 1H), 7.13–7.36 (m, 20H). HR m/z 442.2156 correlates with the chemical formula C₃₀H₂₉NNa within ≤3 ppm.

2.1.4. N-(3,3-Diphenyl)propyl-2-hydroxy-2,2-diphenylacetamide (4d)

Benilic acid (300 mg, 1.31 mmol) and 3,3-diphenylpropylamine (276 mg, 1.31 mmol) were dissolved in DMF (4 mL), and the resulting solution was cooled to 0 °C. After 30 min, HBT (195 mg, 1.44 mmol) and N-methylmorpholine (265 mg, 2.62 mmol) were added, and the resulting mixture was stirred at 0 °C for 1 h, then DCC (300 mg, 1.45 mmol) was added. The resulting mixture was stirred at room temperature overnight, then washed with NaHCO₃ saturated solution and brine, and extracted with EtAc. Organic phase was dried (Na₂S₄), and solvent was removed under reduced pressure. The resulting residue was purified by silica gel column chromatography using n-hexane/EtAc (7/3) as eluent to give the title compound (490 mg, 93%) as a white solid mp 134–135 °C ¹H NMR (CDCl₃) δ 2.32 (m, 2H), 3.32 (m, 2H), 3.84 (t, 1H, 7.9 H), 3.98 (s, 1H), 6.34 (br s, 1H), 6.92–7.43 (m, 20H). HR m/z 444.1920 correlates with the chemical formula C₂₉H₂₇N₂Na within ≤3 ppm.

2.2. Synthesis of compounds 9a–g, 10a–c, and 11a–c

2.2.1. 2-(4-Chlorophenyl)-2-phenyl-N-(3,3-diphenylpropyl)-acetamide (9a)

To a stirred solution of 2-(4-chlorophenyl)-2-phenylacetic acid¹⁶ (500 mg, 2.03 mmol) in dry dichloromethane (10 mL), N,N'-carbonyldiimidazole (658 mg, 4.06 mmol) and 4-dimethylaminopyridine (123 mg, 1.01 mmol) were added at room temperature. After 30 min, 3,3-diphenylpropylamine (0.9 mL, 4.06 mmol) was added. The mixture was stirred at room temperature for 4 h, then solvent was removed under reduced pressure, and the resulting residue was taken up in EtAc and washed with brine. The organic portion was dried (Na₂S₄), the solvent was evaporated and the resulting residue was purified by silica gel column chromatography (hexane/EtAc, 6/4) to give the title compound 9a (709 mg, 80% yield) as a white solid mp 115–120 °C ¹H NMR (CDCl₃) δ 7.40–7.18 (m, 19H) 5.52 (br s, 1H) 4.81 (s, 1H) 3.87 (t, 1H, 7.9 H) 3.29 (s, 2H, 6.7 H) 2.29 (s, 2H, 7.4 H). HR m/z

462.1592 correlates with the chemical formula C₂₉H₂₆NClNa within ≤1 ppm.

2.2.2. 2-(4-Bromophenyl)-2-phenyl-N-(3,3-diphenylpropyl)-acetamide (9b)

Starting from 2-(4-bromophenyl)-2-phenylacetic acid¹⁶ (710 mg, 2.43 mmol), the title compound was obtained as reported for 9a as a white solid in a 87% yield mp 109–115 °C ¹H NMR (CDCl₃) δ 7.49 (d, 2H, 8.4 H) 7.37–7.12 (m, 17H) 5.52 (br s, 1H) 4.78 (s, 1H) 3.89 (t, 1H, 7.8 H) 3.29 (s, 2H, 6.7 H) 2.29 (s, 2H, 7.4 H). HR m/z 506.1102 correlates with the chemical formula C₂₉H₂₆NBrNa within ≤2 ppm.

2.2.3. 2-(4-Methoxyphenyl)-2-phenyl-N-(3,3-diphenylpropyl)-acetamide (9c)

Starting from 2-(4-methoxyphenyl)-2-phenylacetic acid¹⁶ (2 g, 8.54 mmol), the title compound was obtained as reported for 9a as a white solid in a 75% yield mp 100–105 °C ¹H NMR (CDCl₃) δ 7.45–7.18 (m, 17H) 6.89 (d, 2H, 8.6 H) 5.56 (br s, 1H) 4.83 (s, 1H) 3.90–3.83 (m, 4H) 3.28 (s, 2H, 6.7 H) 2.28 (s, 2H, 7.3 H). HR m/z 458.2098 correlates with the chemical formula C₃₀H₂₉N₂Na within ≤1 ppm.

2.2.4. 2-(4-Methylphenyl)-2-phenyl-N-(3,3-diphenylpropyl)-acetamide (9d)

Starting from 2-(4-methylphenyl)-2-phenylacetic acid¹⁶ (2.5 g, 11.05 mmol), the title compound was obtained as reported for 9a as a white solid in a 72% yield mp 125 °C ¹H NMR (CDCl₃) δ 7.38–7.13 (m, 19H) 5.55 (br s, 1H) 4.86 (s, 1H) 3.87 (t, 1H, 7.8 H) 3.27 (s, 2H, 6.7 H) 2.37 (s, 3H), 2.28 (s, 2H, 7.5 H). HR m/z 442.2154 correlates with the chemical formula C₃₀H₂₉NNa within ≤3 ppm.

2.2.5. 2-(4-Nitrophenyl)-2-phenyl-N-(3,3-diphenylpropyl)-acetamide (9e)

Starting from 2-(4-nitrophenyl)-2-phenylacetic acid¹⁶ (2.5 g, 9.72 mmol), the title compound was obtained as reported for 9a as a white solid in a 72% yield mp 125 °C ¹H NMR (CDCl₃) δ 8.20 (d, 1H, 8.9 H), 7.45–7.18 (m, 18H) 5.53 (br s, 1H) 4.83 (s, 1H) 3.88 (t, 1H, 8.0 H) 3.32 (s, 2H, 6.6 H) 2.30 (s, 2H, 7.5 H). HR m/z 473.1803 correlates with the chemical formula C₂₉H₂₆N₃Na within ≤7 ppm.

2.2.6. 2-(4-Hydroxyphenyl)-2-phenyl-N-(3,3-diphenylpropyl)-acetamide (9f)

To a solution of sodium iodide (900 mg, 6.07 mmol) and amide 9c (1.2 g, 2.76 mmol), in CH₃CN (20 mL), chlorotrimethylsilane (0.80 mL, 6.07 mmol) was added at 23 °C. After heating under reflux for 16 h, the reaction was quenched with water and extracted with ether. The organic layer was washed with saturated Na₂S₂O₃ and brine, dried over Na₂S₄, and evaporated. Column chromatography (silica gel, 11 petroleum ether/ether) gave 9f as a white solid in a 60% yield mp 145–150 °C ¹H NMR (CDCl₃) δ 7.39–7.18 (m, 17H) 6.89 (d, 1H, 8.5 H) 6.77 (d, 1H, 8.5 H) 5.56 (br s, 1H) 4.82 (s, 1H) 3.89 (t, 1H, 8.0 H) 3.27 (s, 2H, 6.7 H) 2.28 (s, 2H, 7.3 H). HR m/z 444.1921 correlates with the chemical formula C₂₉H₂₇N₂Na within ≤3 ppm.

2.2.7. 2-(4-Aminophenyl)-2-phenyl-N-(3,3-diphenylpropyl)-acetamide (9g)

A solution of amide 9e (1.5 g, 3.32 mmol), inc dust (2.12 g, 33.2 mmol), absolute ethanol (40 mL), and 37HCl (15 mL) at –10 °C was mixed under reflux for 2 h. After addition of 10 NaH (pH ~10), the organic layer was extracted with ether, washed with brine, dried over Na₂S₄, and evaporated. Column chromatography (silica gel, 11 petroleum ether/ethyl acetate) gave 90% yield of 9g

1.511)0.58 (d, 2H, 6.41)0.65 (d, 2H, 6.41)5.57 (d, s, 1H)4.78 (s, 1H)3.83 (t, 1H, 6.0 H)3.22 (, 2H, 5.0 H) 2.17 (, 2H, 5.0 H). HR m 443.2101 correlates with the chemical formula C₂₉H₂₈N₂Na within ≤2 ppm.

2.2.8. N-(3-Phenyl)propyl-2,2-diphenylacetamide (10a)

Starting from 2,2-diphenylacetic acid (400 mg, 1.89 mmol) and 3-phenylpropylamine (383 mg, 2.83 mmol), the title compound was obtained as reported for 9a as a white solid in a 63% yield mp 135 °C ¹H NMR (CDCl₃) δ 7.39–6.94 (m, 15H), 5.60 (br s, 1H), 4.88 (s, 1H), 3.40 (, 2H, 6.9 H), 2.66 (t, 2H, 7.0 H), 1.89–1.77 (m, 2H). HR m 352.1678 correlates with the chemical formula C₂₃H₂₃NNa within ≤2 ppm.

2.2.9. N-(3-Phenyl)propyl-2-(4-chlorophenyl)-2-phenylacetamide (10b)

Starting from 2-(4-chlorophenyl)-2-phenylacetic acid¹⁶ (620 mg, 2.52 mmol), and 3-phenylpropylamine (681 mg, 5.04 mmol) the title compound was obtained as reported for 9a as a white solid in a 70% yield mp 125 °C ¹H NMR (CDCl₃) δ 7.35–7.05 (m, 14H), 5.57 (br s, 1H), 4.86 (s, 1H), 3.35 (, 2H, 6.7 H), 2.62 (t, 2H, 7.6 H), 1.89–1.80 (m, 2H). HR m 386.1250 correlates with the chemical formula C₂₃H₂₂NCINa within ≤10 ppm.

2.2.10. N-(3-Phenyl)propyl-2-(4-bromophenyl)acetamide (10c)

Starting from 2-(4-bromophenyl)-2-phenylacetic acid¹⁶ (1.85 g, 6.35 mmol) and 3-phenylpropylamine (1.72 g, 12.70 mmol), the title compound was obtained as reported for 9a as a white solid in a 81% yield mp 132 °C ¹H NMR (CDCl₃) δ 7.50–7.10 (m, 14H), 5.57 (br s, 1H), 4.84 (s, 1H), 3.34 (, 2H, 6.7 H), 2.62 (t, 2H, 7.6 H)1.96–1.82 (m, 2H). HR m 430.0765 correlates with the chemical formula C₂₃H₂₂NBrNa within ≤3 ppm.

2.2.11. N-(3,3-Diphenyl)propyl-2-phenylacetamide (11a)

Starting from phenylacetic acid (300 mg, 2.21 mmol) and 3,3-diphenylpropylamine (700 mg, 3.31 mmol), the title compound was obtained as reported for 9a as a white solid in a 85% yield mp 85 °C ¹H NMR (CDCl₃) δ 8.0 (s, 1H), 7.41–7.06 (m, 15H), 4.10 (t, 1H, 7.2 H), 3.44 (s, 2H), 3.33 (, 2H, 6.9 H), 2.19 (, 2H, 7.5 H). HR m 352.1675 correlates with the chemical formula C₂₃H₂₃NNa within ≤2 ppm.

2.2.12. N-(3,3-Diphenyl)propyl-2-(4-chlorophenyl)acetamide (11b)

Starting from 4-chlorophenylacetic acid (500 mg, 2.93 mmol) and 3,3-diphenylpropylamine (1.2 g, 5.86 mmol), the title compound was obtained as reported for 9a as a white solid in a 71% yield mp 107 °C ¹H NMR (CDCl₃) δ 7.37–7.16 (m, 14H), 5.27 (s, 1H), 3.91 (t, 1H, 7.7 H), 3.47 (s, 2H), 3.27 (, 2H, 6.6 H), 2.25 (, 2H, 7.4 H). HR m 386.1247 correlates with the chemical formula C₂₃H₂₂NCINa within ≤10 ppm.

2.2.13. N-(3,3-Diphenyl)propyl-2-(4-bromophenyl)acetamide (11c)

Starting from 4-bromophenylacetic acid (500 mg, 2.32 mmol) and 3,3-diphenylpropylamine (980 mg, 4.64 mmol) the title compound was obtained as reported for 9a as a white solid in a 75% yield mp 110 °C ¹H NMR (CDCl₃) δ 7.50 (d, 2H, 7.4 H), 7.32–7.12 (m, 10H), 6.96 (d, 2H, 7.5 H), 5.33 (br s, 1H), 3.90 (, 1H, 7.5 H), 3.46 (s, 2H)3.24 (, 2H, 6.6 H), 2.25 (, 2H, 7.2 H). HR m 430.0776 correlates with the chemical formula C₂₃H₂₂NBrNa within 1 ppm.

2.3. Binding assay

For CB₁ and CB₂ receptor binding assays, the new compounds were tested using membranes from HEcells transfected with either the human CB₁ or CB₂ receptor and ³H-(–)-cis-3-(2-hydroxy-4-(1,1-dimethylheptyl)-phenyl)-trans-4-(3-hydroxy-propyl)-cyclohexanol (³HCP-55,940) (K_d 0.31 nM for CB₂ and 0.18 nM for CB₁ receptors) as the high affinity ligand as described by the manufacturer (Perkin-Elmer, Italia).²³ Displacement curves were generated by incubating drugs with ³HCP-55,940 (0.084 for CB₂ and 0.14 nM for CB₁ binding assay). In all cases, IC₅₀ values were calculated by applying the Cheng–Prusoff equation to the IC₅₀ values (obtained by GraphPad) for the displacement of the bound radioligand by increasing concentrations of the test compounds.

2.4. cAMP assay

Cyclic AMP assays were performed on intact confluent N18T2 cells plated in six-well dishes and stimulated for 10 min at 37°C with forskolin 1 μM in 400 μl of serum-free Dulbecco's modified Eagles medium containing 20 mM Hepes, 0.1 mg/ml BSA, 0.1 mM 1-methyl-3-isobutylxanthine.²⁴ Cells were treated with vehicle (methanol, 0.1) or compounds (at various concentrations) or IN-55,212 (100 nM) or IN-55,212 plus compound 4b (1 μM). After incubation, 800 μl of ethanol was added, cells were extracted and cyclic AMP was determined by means of a cyclic AMP assay kit (Amersham,), as advised by the manufacturer.

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